Combining machine learning and mechanistic modeling approaches to solve real life problems – assessment of the local tissue binding and its influence on the systemic exposure after topical application of drugs





<u>Sebastian Polak</u> (<u>sebastian.polak@certara.com</u>)^{1,2} Aleksander Mendyk² Nikunjkumar Patel¹

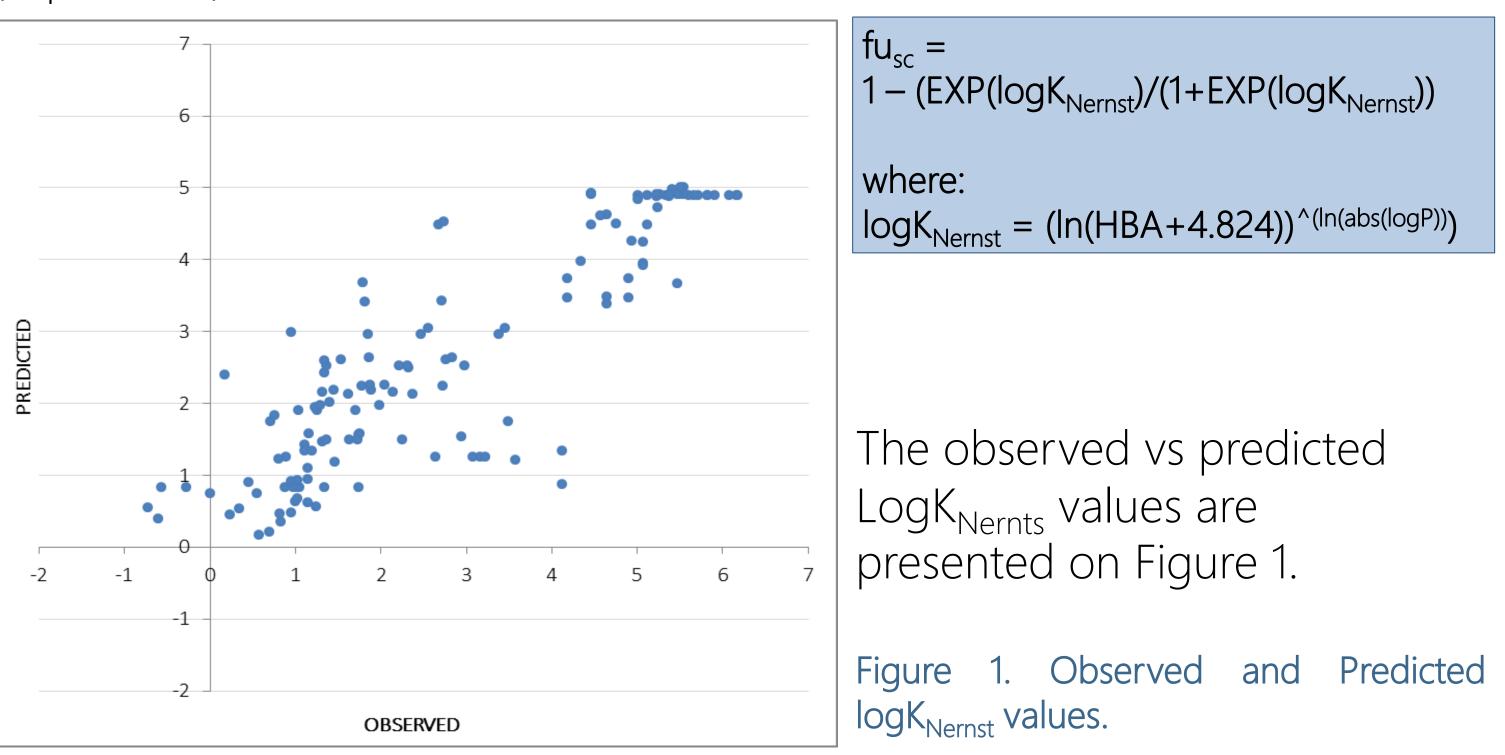
¹Certara UK Ltd, Simcyp Division, Sheffield, UK ²Faculty of Pharmacy, Jagiellonian University Medical College, Kraków, Poland

Introduction

Dermal drug administration can be a preferred route for the delivery of drugs for local or systemic action, with numerous advantages over oral administration. Physiologically Based Pharmacokinetic (PBPK) models have recently gained significant attention in regulatory submissions to quantitatively predict drug-drug interactions (DDIs) and absorption/bioavailability processes [1]. A PBPK approach has a strong potential to help bridge the gap in clinical knowledge in a situation where clinical studies are either difficult or practically infeasible.

Results

The best obtained model for the protein binding in stratum corneum had RMSE = 0.92 and NRMSE = 13.31%. The final equation is presented below (Equation 1).

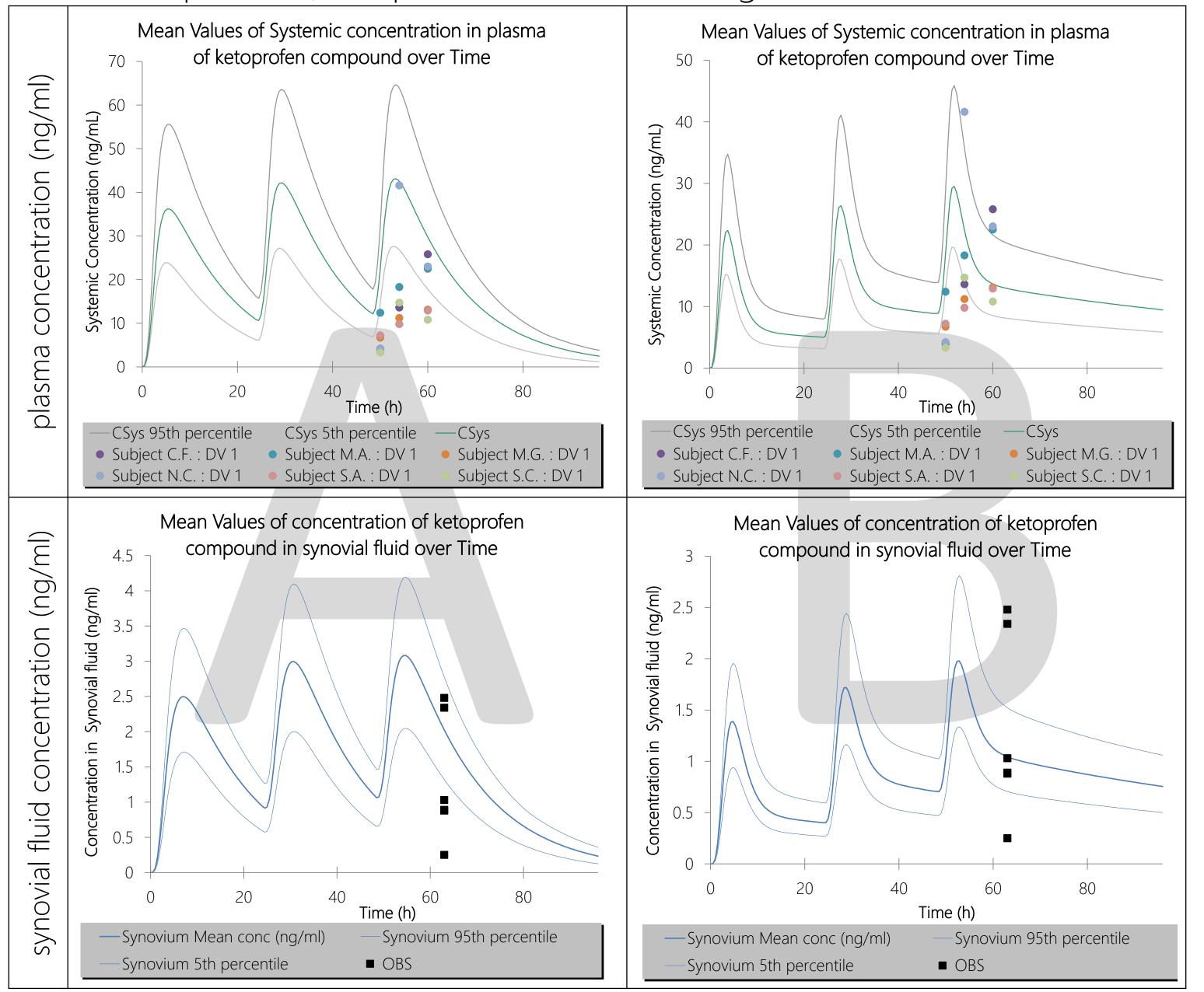


Drug binding in the human tissues is one of the crucial parameters needed for the proper prediction of the drug exposure. Assuming that only unionized and unbound molecule can diffuse through the skin and subcutis tissues, and that the experimental measurement of this parameter is not routinely done, its prediction becomes crucial. In the current study we applied machine learning approach to develop empirical model for the stratum corneum binding. In the next step we utilized the predicted values to assess their influence on the dermally applied model drug (ketoprofen) local and systemic exposure predicted with the use of mechanistic PBPK model (Simcyp MPML MechDermA model) [1].

Methods

Binding coefficients to different keratins, namely, bovine hoof and horn, human delipidized callus, human delipidized stratum corneum (SC), human nail, human hair, and sheep wool collected by Hansen and colleagues from the literature, were utilized for the modeling purposes [4]. The modelled endpoint was Nernst binding coefficient K_{Nernst} (log K_{Nernst}). The model was developed using data-driven approach based on the gathered data set and bioinspired heuristic algorithm so-called genetic programming (GP), based on simple physico-chemical parameters (logP, HBA). a machine learning system belonging to the evolutionary GP İS computation (EC) systems, where solution is represented as an artificial chromosome and based on available data developed iteratively by means of genetic operators (mutation, crossing-over, etc.) performed on the population of chromosomes towards the optimum state. In this work we used a variant of GP called symbolic regression where the artificial chromosome represents mathematical equation, both its structure and coefficients.

MechDermA model simulation results for the Ballerini study for two scenarios: A) $fu_{sc} = 1$ (lack of binding assumed), B) $fu_{sc} = 0.093$ (predicted with the Equation 1) are presented below on Figure 2.



R statistical environment [6] was used to perform all calculations with GP implementation in rgp module [7] extended by Pacławski et al. [8]. The model quality was assessed with the use of root mean squared error (RMSE) and normalized RMSE (NRMSE). The predicted fusc value for ketoprofen (0.093) was further applied to the MPML MechDermA model, and the systemic concentration predictions for fusc = 1, and the predicted value were compared.

Table 1a, b. Ketoprofen ADME / skin permeation parameters.

а	Group		Param	neter	Va	lue [unit]	Source	Ballerini et al.
b			MW		254.285		[2]	reported plasma
			logP		3.12		[3]	and synovial fluid
	Phys-ch	iem	Туре		monoprotic acid		[2]	concentration
			рКа		4.45		[2]	after topical
	Binding		B/P partition ratio		1.1		[4]	application of
			fu _{plasma}		0.01		[3]	ketoprofen gel
	Distribution		Vss		0.1	D.132 [L/kg]; CV=17.2% [5]		[9]. 10cm long
	Elimination		CLIV		5.16 [L/h]; CV=17.4%		[5]	strip containing
	Group Parame		meter	Value [unit]		Source		70-80 mg of
	•	Kp _{sc_lip:vehicle}		162.5451		predicted – Hansen 2013		ketoprofen was
		Kp _{sc:ve}		6.9144		predicted – Kretsos 2008		applied on the
		Kp _{derm}	iis:ve	1		assumed	8	knee (assumed
		Kp _{sebul}	m:vehicle	1643.341		predicted – Valiveti et al. 2008		
	L L C	Kp _{skin:b}	blood	2.67544		predicted – Shatkin&Brown 1991		area 400 cm ²).
	Absorption	Kp _{sebul}	m:ve	0.0989		calculated – Kp _{lip:vehicle} /Kp _{sebum:vehicle}		Plasma and
	pso	D _{sc_lip}		3.392E-06 [cm ² /h]		predicted – Mitragotri et al. 2003		
		D _{ve}		0.000311 [cm ² /h]		predicted –Bunge&Cleek		synovial fluid concentrations were measured.
		D _{dermis}	5	0.000311 [cm ² /h]		predicted –Kretsos et al. 2008		
		D _{sebum}	1	0.000653 [cm ² /h]		predicted – Johnson et al. 1996		
		fu sc		0.0934		predicted – Polak et al. 2016		
		f _{ni, corne}	eocytes	1		assumed		

Figure 2. MechDermA model simulation results for two tested scenarios.

Discussion and Conclusions

The current study presents the problem solving approach based on combining machine learning and mechanistic modeling. Drug absorption after dermal application was used as an exemplary problem. As only the free, unbound drug can permeate and partition between skin layers, the knowledge about binding to the SC structures allows proper

parametrization of the model. This value is although not commonly measured, and the QSAR model utilizing simple physico-chemical data.

Ketoprofen plasma exposure after topical application was simulated with the assumption that the binding in SC is negligible ($fu_{sc}=1$). The model overpredicted the plasma concentration. Based on the ketoprofen specific phys-chem data the fu_{sc} value was calculated (fu_{sc}=0.093) and applied to the MechDermA model. The simulated exposure closely mimicked the clinically observed plasma concentrations.

References

1) Martins F, et al. 2017 GRC - Barrier Function of Mammalian Skin, Waterville Valley, NH., 2) https://pubchem.ncbi.nlm.nih.gov/compound/3825; 3) https://www.drugbank.ca/drugs/DB01009; 4) Fura A, et al. Biopharm Drug Dispos. 2008;29(8):455-68.; 5) Debruyne D, et al. Clin Pharmacokinet. 1987;12(3):214-21.; 6) http://www.R-project.org; 7) Flasch O. 2014; https://CRAN.R-project.org/package=rgp; 8) Pacławski A. http://sourceforge.net/projects/rscriptsmultivariate/files/rgp/. 9) Ballerini R. et al. Int J Clin Pharmacol Res. 1986;6(1):69-72.

ACoP9, October 6th – 10th, 2018 San Diego, USA